

Copper-Catalyzed C(sp³)–OH Cleavage with Concomitant C–C Coupling: Synthesis of 3-Substituted Isoindolinones

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Supporting Information



ABSTRACT: Copper(II) trifluoromethanesulfonate $(Cu(OTf)_2)$ efficiently catalyzes the C–C coupling of 3-hydoxyisoindolinones with a variety of aryl-, heteroaryl-, and alkenylboronic acids to furnish C(3) aryl-, heteroaryl-, and alkenyl-substituted isoindolinones. The coupling reactions work smoothly in 1,2-dicholoroethane (DCE) reflux, to effect both inter- and intramolecular versions. This is the first report on C(sp³)–OH cleavage with concomitant C–C coupling. The photolabile 2-nitrobenzyl protecting group is most appropriate for promotion of the coupling reaction and for deprotection. The tetracyclic ring motif of the alkaloid neuvamine was prepared by applying the newly developed copper-catalyzed C–C coupling.

INTRODUCTION

Unlike indole, its isomer, isoindole (2H-isoindole), is not a common structural element in natural products.¹ However, bioactive molecules built around the isoindole structure are considered privileged due to their multifarious medicinal properties.² Among isoindoles, the C(3)-substituted isoindolinones occur as a partial structure in a few alkaloids. Representative examples of such isoindole alkaloids are nuevamine (1), the first known isoindoloisoquinoline alkaloid, and lennoxamine (2), an isoindolobenzazepine alkaloid, both of which have been isolated from Berberis darwinii (Figure 1).³ Pestalachloride A (3), an antifungal alkaloid isolated from the endophytic fungus Pestalotiopsis adust,⁴ and taliscanine (4), an anti-Parkinson alkaloid isolated from the rhizomes of Aristolochia taliscana,⁵ are other examples of alkaloids with a C(3)-substituted isoindole structure (Figure 1). In addition to the medicinally important alkaloids listed above, some central nervous system (CNS) active drug candidates such as (S)-pazinaclone (5) and (R)-PD 172939 $(6)^6$ possess a C(3)-substituted isoindole structure (Figure 1). Apart from these two drug candidates, the C(3)substituted isoindolinones exhibit varied biological activities that include antipsychotic,⁷ antihypertensive,⁸ antiulcer,⁹ and anxiolytic¹⁰ properties. In view of the importance of isoindolinones, there have been great synthetic efforts toward these coveted structures.¹¹

The C(3)-substituted isoindolinones **9** can be viewed as derivatives of phthalimide. Generally they have been synthesized from their hydroxy counterparts 7 by S_N1 substitution at the hydroxy carbon with electron-rich aromatic compounds (carbon nucleophiles; Scheme 1). Due to the poor leaving ability of the hydroxy group, strong protic acids such as triflic acid,¹² trifluoroacetic acid,¹³ concentrated H₂SO₄,¹⁴ and

concentrated HCl^{15} or Lewis acids such as $Bi(OTf)_{3}^{16}$, $Sn(NTf_2)_{4}^{17}$ Ir–Sn₃ bimetallic complexes, ¹⁸ and gold catalysts¹⁹ have been employed to generate iminium ion 8, for quenching with electron-rich aromatic compounds or nucleophiles. Such reactions are generally conducted on 7, having a nitrogen protecting group (PG). Indeed, most of the researchers to date have taken recourse to robust benzyl as the nitrogen PG, but the benzyl group is difficult to remove without destroying the isoindolinone ring. Overall, existing methods are beleaguered with several drawbacks such as the requirement of (i) concentrated protic acids or Lewis acids, (ii) electron-rich aromatic compounds to quench the acyl minium ion, and (iii) a protecting group that cannot be removed without affecting the isoindolinone ring. Thus, there is a need to discover conditions for facile replacement of the hydroxyl group in 7 with electrondeficient aromatic, heteroaromatic, or alkenyl groups through C-C coupling without going through acyliminium ion 8. Furthermore, there is a need for a PG on nitrogen of the isoindolinone ring, which can be removed under non-hydrogenating, neutral, and milder reaction conditions. Toward this goal, we resolved to develop suitable copper-catalyzed C-C coupling reactions and suitable PGs for the synthesis of C(3)substituted isoindolinones.

In recent years, organic synthesis involving transition-metalcatalyzed C–C coupling, via substitution of halides or its congeners involving organometallic intermediates, has been responsible for a paradigm shift in synthetic planning from classical acid–base to coupling pathways.²⁰ Although palladium catalysts have been in the forefront of the emergence of C–C

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Article



Figure 1. Examples of naturally occurring (1-4) and biologically active (5 and 6) C(3)-substituted isoindolinones.

Scheme 1. Existing Method for Substitution of the OH Group in 7 To Form 9 and the Proposed Method for Replacement of the OH Group in 7 with Aryl Groups through C–C Coupling



bond coupling reactions, the high cost and extreme toxicity of the metals, even in trace quantities, forced researchers to look for alternatives.²¹ In this quest, copper catalysts have emerged as viable alternatives. Advantages of copper catalysts include (i) variable oxidation states of copper (+1, +2, and +3), (ii) solubility of many copper salts in organic solvents, (iii) low cost, benchtop availability and air stability, and (iv) environmental and biological compatibility.²² Of late, copper-catalyzed crosscoupling reactions have become a practical choice for building C-O, C-S, and C-N bonds (for example, Chan-Lam--Evans coupling).²³ The Cu-catalyzed C-C coupling reactions, however, have yet to become well established for C-C coupling, possibly due to low yields and harsh reaction conditions (e.g., Ulmann coupling).²⁴ With the advent of organoboron reagents, copper-catalyzed C-C coupling reactions are becoming feasible on a variety of substrates.²⁵ Organoboranes are stable, exhibit higher functional-group tolerance, and are commercially available or easy to prepare.²⁶ We have recently demonstrated facile copper-catalyzed $C(sp^3)-C(sp^2)$ cross-coupling reactions of organoboron reagents with 4H-chromenes having a C(4) SMe group to furnish a variety of 4-aryl-4H-chromenes.²⁷ In a continuation of this study, we present herein the coppercatalyzed intermolecular/intramolecular coupling reaction of isoindolinol 7 with aryl-, heteroaryl-, and alkenylboronic acids for the facile synthesis of C(3)-substituted isoindolinones (Scheme 1). Although some examples of Pd(II)- or Ni(0)catalyzed cross-coupling of organoboron reagents with activated benzylic hydroxyl are known,²⁸ ours is the first report on such cross coupling involving a free benzylic hydroxyl group, an arylboronic acid, and a copper catalyst.

RESULTS AND DISCUSSION

Our initial efforts were directed toward unearthing a suitable copper catalyst and reaction conditions for coupling of *N*-benzyl-protected 3-hydroxyisoindolin-1-one (7**a**) and phenyl-boronic acid (12**a**) to furnish 2-benzyl-3-phenylisoindolin-1-one (10**a**) (Table 1). Following our experience,²⁷ as a first attempt, we employed reflux of 10 mol % of Cu(OAc)₂·H₂O in dichloromethane (DCM), but the reaction provided 10**a** in less than 5% yield. The yield rose to 45% when the reaction was conducted in higher boiling dichloroethane (DCE) reflux

(entry 1). The yield gradually rose to a plateau of about 85% with increased catalytic loading (entries 2 and 3). Alternate Cu(II) catalysts such as $Cu(CF_3COO)_2 \cdot H_2O_1$, $Cu(acac)_2$ CuCl₂·2H₂O, CuBr₂, CuSO₄·5H₂O, and CuO (entries 4–9) did not provide the desired 10a in better yield. However, in the genre of Cu(II) catalysts, 10 mol % of copper(II) trifluoromethanesulfonate $(Cu(OTf)_2)$ provided the best yield (86%) after chromatographic purification of the product (entry 10). A smaller amount of catalytic loading (5 mol %) was not sufficient to complete the transformation in a reasonable time (entry 12), and a higher amount of catalytic loading (20 mol %) did not improve the yield further. Moreover, we made certain that $Cu(OTf)_2$ catalysis was not due to a minor triflic acid impurity, by conducting the reaction in the presence of sodium carbonate as the buffering agent (entry 11). In reality, the yield perceptibly rose to 92% in the presence of 0.5 equiv of Na_2CO_3 , indicating that a minor amount of TfOH, which is inevitable in commercial $Cu(OTf)_2$, may actually impede the reaction. Furthermore, a run with 10 mol % of TfOH alone was conducted to convincingly rule out the possibility of TfOH being responsible for the reaction as a Brønsted acid. As anticipated, the reaction did not proceed to provide **10a** (entry 13). Copper(I) salts such as CuCl, CuBr, CuI, and Cu_2O or stable copper(I) complexes such as CuBrSMe₂, Cu(PPh₃)₃Br, copper(I) thiophene-2-carboxylate (CuTC), and Cu(I) 3-methylsalicylate (CuMeSal) did not promote the coupling (entries 14–21), indicating that copper(I) species may not be involved in the reaction. The low yield obtained in the cases of CuCl, CuBr, CuTC, and CuMeSal can be attributed to Cu(II) impurities or a facile switch of Cu(I) to Cu(II). Notably, there was no reaction when a palladium(II) catalyst $(Pd(OAc)_{2})$ entry 22) or palladium(0) catalyst (Pd₂(dba)₃, entry 23) was employed in the presence or absence of a base. To evaluate if $Cu(OTf)_2$ catalysis is due to its Lewis acidic nature, we conducted the reaction in the presence of catalytic amounts of similar borderline Lewis acids^{29⁻} such as Sc(OTf)₃, Fe(OTf)₃, Zn(OTf)₂, and Yb(OTf)₃, but the reactions did not work (entries 24-27).

As an alternative to DCE, we screened some common solvents and changed the reaction parameters to decipher the best conditions. Of the alternate solvents investigated such as toluene (42%), tetrachloroethane (TCE; 76%), acetonitrile (28%), and

Table 1. Optimization Conditions of Coupling ReactionsConducted in DCE Reflux in the Presence of DifferentCatalysts



dioxane (48%), none of them worked as well as DCE (92%). The transformation worked best under an atmosphere of nitrogen. Under an oxygen atmosphere the yield of **10a** was only 16%, indicating that the reaction does not go through oxygenmediated catalyst regeneration. Surprisingly, metal complexing and organic solvent solubilizing ligands such as PPh₃ (34%) and phenanthroline (42%) actually decreased the yield of **10a**.

Next, we looked into the reactivity of two derivatives of phenylboronic acids, namely phenylboronic acid pinacol ester (12b) and potassium phenyltrifluoroborate (12c), toward the coupling reaction, since both of the reagents have been employed previously in place of phenylboronic acid with concomitant advantages (Scheme 2).³⁰ However, we found that in such runs there is not much difference in the yield of 10a over the reaction when phenylboronic acid was employed. As a result, because of their easy availability, we chose to go ahead with phenylboronic acids for further studies.

With the optimized conditions in hand, we evaluated the generality of 3-arylisoindolinone synthesis and outcome of the reaction as a consequence of the electron density of the aromatic ring (Scheme 3). We conducted coupling between 2-benzyl-3-hydroxyisoindolinone (7a) and three more arylboronic acids:

Scheme 2. Scope of Cu-Catalyzed Cross-Coupling of Arylboronic Acid Derivatives with 2-Benzyl-3-hydroxyisoindolinone (7a)



Scheme 3. Scope of Cu-Catalyzed Cross-Coupling of Different Arylboronic Acids with 2-Benzyl-3-hydroxyisoindolinone (7a)



namely, 4-trifluoromethylphenylboronic acid (12d), having a highly electron withdrawing C4-CF₃ group, 4-methoxyphenylboronic acid (12e), having a highly electron donating C4-OMe group, and furyl-2-boronic acid (12f), in which the boronic acid is on an electron-rich heteroaromatic ring. Each reaction provided the corresponding 3-arylisoindolinones 10b-d in good to excellent yield. Among the three boronic acids 12d-f, the reaction with furyl-2-boronic acid (12f) provided the lowest yield (78%) and the reaction took a longer time (12 h), reflecting the sluggish nature of the reactant in coupling reactions.³¹ The isoindolinone 10a was characterized by spectral (IR, ¹H NMR, ¹³C NMR, and DEPT-135) and analytical data. A singlet at about δ 5.0 ppm assignable to C(3)H in the ¹H NMR spectrum of 2-benzyl-3-phenylisoindolin-1-one (10a) is the diagnostic signal. Spectral and analytical data of 10b-d compared well with those of 10a.

Having stabilized reaction conditions for the coppercatalyzed cross coupling of 7a and aryl boronic acids 12, it was our next endeavor to remove the benzyl PG in 10a so that the isoindolinone motif would be exposed. As anticipated, selective reductive removal of the PG proved to be difficult, as there are two benzylic positions in 10a, both of which were becoming reductively cleaved under hydrogenation conditions, resulting in destruction of the isoindolinone motif. As an alternative, we placed a 4-methoxybenzyl group (PMB) on the nitrogen in 7b with the intent of selectively removing it under oxidative conditions (Scheme 4).32 Regrettably, the coupling reaction between 7b and phenylboronic acid (12a) was not efficient. Surprisingly, when palladium-sensitive allyl species 7c and propargyl species $7d^{33}$ were placed on nitrogen, there was no reaction to provide the anticipated 10f,g respectively. It is possible that 7c,d and $Cu(OTf)_2$ combine to provide stable complexes. Gratifyingly, when we employed the photolabile 2-nitrobenzyl (NB) PG^{34} on the isoindolinone motif (7e), the coupling worked best to provide product 10h in excellent yield within 3 h.

Scheme 4. Scope of Cu-Catalyzed Cross-Coupling in the Presence of Different Protecting Groups (PGs)



To understand the reaction mechanism of $C(sp^3)-C(sp^2)$ coupling and to rule out the possibilities of nucleophilic substitution, we conducted two reactions on 3-hydroxyisoindolinone (7a) in THF: one with PhMgBr, a hard nucleophile, and the other with 2(PhMgBr)CuI,³⁵ a soft nucleophile. Neither reaction provided the substitution product **10a**, ruling out the possibility of a nucleophilic substitution. As noted in Table 1 (entries 10 and 11), a TfOH impurity in Cu(OTf)₂ or TfOH (entry 13) impedes the reaction by engineering toward the generation of *N*-acyliminium ion intermediates, instead of channeling the reactant 7 toward coupling pathways. On the basis of the evidence accumulated so far, a possible mechanism could be depicted as shown in Scheme 5. The first and

Scheme 5. Plausible Mechanism for Copper-Mediated Formation of 3-Substituted Isoindolinones 10



important step is the insertion of copper into a fragile carbonboron bond to provide the reactive intermediate PhCu(OTf) (13).³⁶ The intermediate 13 then enters into the catalytic cycle to react with 3-hydroxyisoindolinone (7) to provide the intermediate 14. Crucial C-C coupling with concomitant C-O bond cleavage then takes place on 14 to provide the product 10 and copper(II) species 15. The reaction of 15 with arylboronic acid 12 regenerates 13 and stable boric acid. Throughout the catalytic cycle copper remains in the oxidation state of +2. Interaction of copper(II) species with the C(3) hydroxy group in the intermediate 14 could reflect its Lewis acid characteristics. The driving force for the coupling reaction is the formation of a stable C–C bond in 10 and a Cu–O bond in 15, at the cost of Ar-Cu and C-OH bonds. Although metal salt solubilizing ligands such as PPh₃ and phenanthroline helped the dissolution of $Cu(OTf)_2$ in DCE, the decreased yield of the desired product 10 (vide supra) indicates that ligand-bound copper catalyst is sterically hindered to allow the formation of 14.

To demonstrate the scope of the coupling reaction, the arylboronic acids 12d-p were reacted with 3-hydroxy-2-(2-nitrobenzyl)isoindolin-1-one (7e) under the optimized conditions to give the 15 2-(2-nitrobenzyl)-3-arylisoindolin-1-ones

Table 2. Scope of Cu-Catalyzed Cross-Coupling
of 2-Nitrobenzyl-Protected 3-Hydroxyisoindolinone 7e
with Different Aryl/Heteroaryl/Alkenylboronic Acids

7	O N [−] NB ₊ RB(OH OH Ze 12d-r	Cu(O) Na ₂ Ci DCE, 4-10 b	Tf) ₂ (10 mol' O ₃ (0.5 equi reflux n, 63-92%	%) v)	0 N-NB R 10i-w
entry	R	substrate	product	time (h)	yield (%)
1	$4-CF_3C_6H_4$	12d	10i	4	92
2	4-MeOC ₆ H ₄	12e	10j	10	77
3	2-furyl	12f	10k	6	79
4	$4-FC_6H_4$	12g	101	4	91
5	3,5-F ₂ C ₆ H ₃	12h	10m	5	81
6	4-ClC ₆ H ₄	12i	10n	6	81
7	3-ClC ₆ H ₄	12j	10o	6	88
8	$4-BrC_6H_4$	12k	10p	8	85
9	4-MeC ₆ H ₄	121	10q	8	89
10	3-MeC ₆ H ₄	12m	10r	10	86
11	$2,3-(OMe)_2C_6H_3$	12n	10s	6	79
12	$2,5-(OMe)_2C_6H_3$	120	10t	6	81
13	$2,6-(OMe)_2C_6H_3$	12p	10u	10	74
14	n-hex-1-enyl	12q	10v	4	63
15	styryl	12r	10w	6	84

10i-u (Table 2). The arylboronic acids 12d-p were selected with a view toward their structural diversity and potential binding to biological targets. High efficiency of cross-coupling was observed regardless of the presence of strongly electron withdrawing (12d,g,h to 10i,l,m), mildly electron withdrawing (12i-k to 10n-p), strongly electron donating (12e to 10j), or mildly electron donating (12l,m to 10q,r) nature of the substitution in the aryl ring. Notably, the cross-coupling reaction worked well even with ortho-substituted boronic acids (12n-p to 10s-u), indicating that the cross-coupling is not highly sensitive to the steric bulk on the aryl ring. A boronic acid on an electron-rich heterocyclic ring, namely, furyl-2boronic acid (12f), participated in the coupling with 7e to furnish isoindolinone 10k. However, the coupling did not work with thiophene-2-boronic acid, pyridene-3-boronic acid, and quinolone-3-boronic acid, indicating that the reaction is sensitive to boronic acids that contain coordinating sites. The copper-mediated cross-coupling between 3-hydroxyisoindolin-1-one 7e and alkenylboronic acids 12q,r to furnish alkenyl substituted isoindolinones 10v,w took place without any difficulty.

After we demonstrated the feasibility of synthesizing different C(3)-substituted isoindolinones **10h**–w with NB as the N-protecting group, our next task was to cleave photolabile NB, so that the free isoindole moiety becomes exposed. We irradiated dilute (10^{-2} M) CH₃CN/H₂O (1/1) solutions of four selected isoindolinones, namely **10h**,j,l,o, with 4 × 3 μ W

LED lamps emitting at 370 nm (see the Supporting Information for an image of the reactor built in-house).³⁷ The photochemical cleavage reaction was clean and took place within 4 h to provide deprotected products 11a-d in excellent yield (Scheme 6). The

Scheme 6. Cleavage of NB in the Presence of UV Light (370 nm)



photocleavage was facile on substrates having an aryl ring with electron-donating (10j) or electron-withdrawing (10l,o) substituents. We found that the LED lamps as a source of 370 nm light for NB deprotection is far superior in comparison to conventional high-pressure mercury vapor lamps, as LED lamps do not generate heat and a filter to cut off unwanted light is not required.³⁸

Finally, we attempted the synthesis of the tetracyclic lactam system 19 of neuvamine (1) from arylboronic acid 17 to demonstrate an intramolecular version of our newly developed $Cu(OTf)_2$ -catalyzed C–C coupling reaction (Scheme 7). The starting phthalimide derivative³⁹ 16 was prepared from the corresponding amine⁴⁰ and phthalic anhydride in refluxing toluene. The aryl bromide 16 was converted into the boronic acid derivative 17 by a palladium-catalyzed reaction with bis(pinacolato)diboron.⁴¹ Controlled reduction of one of the carbonyl groups in 17 with sodium borohydride provided isoindolin-3-ol 18, which on treatment with $Cu(OTf)_2$ under our optimized reaction conditions provided the neuvamine framework 19 in excellent yield.

CONCLUSION

In summary, we have described facile $Cu(OTf)_2$ -catalyzed $C(sp^3)-C(sp^2)$ coupling involving 3-hydoxyisoindolinones and a variety of aryl-, heteroaryl-, and alkenylboronic acids to efficiently furnish C(3) aryl-, heteroaryl-, and alkenyl-substituted isoindolinones. This is the first report on copper-catalyzed $C(sp^3)-OH$ cleavage with concomitant C-C coupling. In this way, we demonstrated facile substitution of the OH group in 3-hydoxyisoindolinones with electron-rich and electron-deficient aryl and alkenyl groups with equal facility. We have shown that

photolabile 2-nitrobenzyl is the best N-protecting group and DCE reflux is the best medium for the transformation. We demonstrated both inter- and intramolecular versions through the synthesis of 23 C(3)-substituted isoindolinones and the tetracyclic ring motif of the alkaloid neuvamine.

EXPERIMENTAL SECTION

General Experimental Methods. The progress of all reactions was monitored by TLC using a hexanes (60-80 °C boiling mixture)/ ethyl acetate mixture as eluent. Column chromatography was performed on silica gel (100-200 mesh) using increasing percentages of ethyl acetate in hexanes. ¹H NMR (400 MHz), ¹³C NMR (100 MHz), and DEPT-135 spectra were recorded for CDCl₃, CDCl₃/CCl₄ (1/1), or DMSO-d₆ solutions on a 400 MHz spectrometer with TMS as internal standard. Coupling constants J are given in Hz. IR spectra were recorded as KBr pellets on a FT-IR spectrometer. Highresolution mass spectra were recorded on a quadrupole time of flight (QTOF) mass spectrometer using the electrospray ionization mode. The X-ray diffraction measurements were carried out at 298 K on a diffractometer equipped with a graphite monochromator and a Mo K α fine-focus sealed tube ($\lambda = 0.71073$ Å). Organic solvents were dried by standard methods. The catalyst $Pd_2(dba)_3$ and the alkenylboronic acids were prepared according to literature procedures.⁴¹ Lightmediated deprotection of the NB group to generate isoindolinones was carried out using a home-built reactor having four UV-LED $(3 \ \mu W)$ lamps with an emission maximum at 370 nm (see the Supporting Information for the photograph). The maximum intensity of the LED bulbs was determined using a UV-visible spectrometer capable of intensity and wavelength characterization having a resolution of 0.23 nm in the range 200-1100 nm (see Figure 3 in the Supporting Information).

General Procedure for Synthesis of 2-Benzyl-3-arylisoindolin-1-ones 10a-e: Synthesis of 2-Benzyl-3-phenylisoindolinone (10a). An oven-dried 25 mL two-neck round-bottom flask connected to a Schlenk line through a condenser was charged with phenylboronic acid (12a; 60 mg, 0.5 mmol), Cu(OTf)2 (18 mg, 0.05 mmol), and Na2CO3 (27 mg, 0.25 mmol). The flask was sealed with a rubber septum, evacuated under vacuum, and purged three times with nitrogen gas. Anhydrous dichloroethane (DCE, 2 mL) was added through a syringe, and the contents were stirred for 10 min. 2-Benzyl-3-hydroxyisoindolin-1-one (7a; 120 mg, 0.5 mmol) in DCE (4 mL) was next added at room temperature (30 °C) over 5 min. The resulting reaction mixture was refluxed for 4 h while it was periodically checked by TLC for completion of the reaction. The reaction mixture was then extracted with dichloromethane (DCM, 2×20 mL). The organic layer was washed with water $(2 \times 20 \text{ mL})$ and brine $(1 \times 20 \text{ mL})$ 10 mL) followed by removal of DCM under reduced pressure. The crude product was subjected to column chromatography (silica gel, gradient elution with increasing volume of EtOAc in hexanes) to yield 2-benzyl-3-phenyl-isoindolinone (10a).

Scheme 7. Synthesis of Neuvamine-like Molecule 19 via a Cu-Catalyzed Intramolecular Coupling Reaction



2-Benzyl-3-phenylisoindolin-1-one (10a):



colorless solid (143 mg, 92% yield); mp 135 °C (reported 136 °C); IR (KBr, cm⁻¹) 3030, 2920, 1695, 1613, 1494, 1464, 1399, 1291, 1075, 1026, 978, 761, 738, 701; ¹H NMR (400 MHz, CDCl₃) δ 7.94 (d, *J* = 6.6 Hz, 11H), 7.50–7.39 (m, 2H), 7.39–7.02 (m, 1H), 5.40 (d, *J* = 14.8 Hz, 1H), 5.24 (s, 1H), 3.73 (d, *J* = 14.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 168.6 (C), 146.4 (C), 137.1 (C), 136.8 (C), 131.9 (CH), 131.4 (C), 129.2 (CH), 128.8 (2 × CH), 128.5 (CH), 128.4 (CH), 127.8 (CH), 127.6 (CH), 123.8 (CH), 123.2 (CH), 63.6 (CH), 43.9 (CH₂); HRMS (ESI) calcd for C₂₁H₁₇NO (M + H) 300.1382, found 300.1382.

2-Benzyl-3-(4-(trifluoromethyl)phenyl)isoindolin-1-one (10b):



colorless solid (167 mg, 88% yield); mp 115 °C; IR (KBr, cm⁻¹) 3064, 3033, 2926, 2861, 1702, 1618, 1494, 1467, 1398, 1327, 1119, 1068, 891, 848, 795, 737, 700, 609; ¹H NMR (400 MHz, CDCl₃/CCl₄, 1/1) δ 7.95 (d, *J* = 7.1 Hz, 1H), 7.61 (d, *J* = 7.8 Hz, 2H), 7.52–7.37 (m, 2H), 7.28–7.06 (m, 8H), 5.40 (d, *J* = 14.9 Hz, 1H), 5.27 (s, 1H), 3.72 (d, *J* = 14.9 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) 168.4 (C), 145.6 (C), 141.3 (C), 136.8 (C), 132.2 (CH), 131.4 (C), 131.2 (q, *J* = 32 Hz, C), 128.9 (CH), 128.8 (CH), 128.5 (CH), 128.2 (CH), 127.9 (CH), 126.3 (q, *J* = 4 Hz, CH), 125.3 (C), 124.2 (CH), 123.1 (CH), 63.0 (CH), 44.1 (CH₂); HRMS (ESI) calcd for C₂₂H₁₆F₃NONa (M + Na) 390.1082, found 390.1085.

2-Benzyl-3-(4-methoxyphenyl)isoindolin-1-one (10c):



colorless solid (136 mg, 83% yield); mp 124 °C (reported 124 °C); IR (KBr, cm⁻¹) 3062, 2927, 2872, 1694, 1611, 1527, 1468, 1400, 1344, 1305, 1142, 1094, 858, 790, 738, 702; ¹H NMR (400 MHz, CDCl₃) δ 7.86–7.77 (m, 1H), 7.35–7.24 (m, 2H), 7.16–7.11 (m, 3H), 7.06 (d, *J* = 6.6 Hz, 2H), 6.98–6.92 (m, 1H), 6.88–6.69 (m, 4H), 5.25 (d, *J* = 14.8 Hz, 1H), 5.08 (s, 1H), 3.66 (d, *J* = 3.8 Hz, 3H), 3.61 (d, *J* = 14.9 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 168.3 (C), 159.8(C), 146.6 (C), 137.1 (C), 131.8 (CH), 131.3 (C), 129.0 (CH), 128.6 (CH), 128.3 (CH), 128.2 (CH), 127.5 (CH), 123.5 (CH), 123.1 (CH), 114.4 (CH), 63.0 (CH), 55.2 (CH₃), 43.6 (CH₂); HRMS (ESI) calcd for C₂₂H₂₀NO₂ (M + H) 330.1488, found 330.1486.

2-Benzyl-3-(furan-2-yl)isoindolin-1-one (10d):



colorless solid (106 mg, 78% yield); mp 108 °C; IR (KBr, cm⁻¹) 1693, 1592, 1567, 1529, 1305, 1195, 1072, 743, 708; ¹H NMR (400 MHz, CDCl₃) δ 8.00–7.86 (m, 1H), 7.54–7.45 (m, 2H), 7.30–7.19 (m, 6H), 6.29–6.36 (m, 2H), 5.40 (s, 1H), 5.32 (d, *J* = 14.9 Hz, 1H), 3.94 (d, *J* = 14.9 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 168.0 (C), 149.1 (C), 143.5 (CH), 143.0 (C), 137.0 (C), 131.8 (CH), 128.79

(CH), 128.72 (CH), 128.3 (CH), 127.5 (CH), 123.8 (CH), 123.0 (CH), 110.5 (CH), 110.0 (CH), 57.0 (CH), 44.2 (CH₂); HRMS (ESI) calcd for $C_{19}H_{15}NO_2$ (M + H) 290.1175, found 290.1776. 2-(4-Methoxybenzyl)-3-phenylisoindolin-1-one (**10e**):



colorless solid (88 mg, 54% yield); mp 132 °C; IR (KBr, cm⁻¹) 3033, 2929, 2835, 1691, 1612, 1512, 1464, 1399, 1246, 1176, 1033, 817, 737, 699; ¹H NMR (400 MHz, CDCl₃) δ 7.93 (dd, *J* = 6.0, 2.2 Hz, 1H), 7.51–7.30 (m, 5H), 7.17–7.00 (m, 5H), 6.82 (d, *J* = 8.7 Hz, 2H), 5.34 (d, *J* = 14.7 Hz, 1H), 5.22 (s, 1H), 3.77 (s, 3H), 3.68 (d, *J* = 14.7 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 168.5 (C), 159.1 (C), 146.4, 136.9 (C), 131.8 (CH), 127.8 (CH), 129.27 (C), 129.21 (CH), 128.7 (CH), 128.3 (CH), 55.3 (CH₃), 43.2 (CH₂); HRMS (ESI) calcd for C₂₂H₂₀NO₂ (M + H) 330.1488, found 330.1487.

Synthesis of 3-Hydroxy-2-(2-nitrobenzyl)isoindolin-1-one (7e).



To a stirred solution of 2-(2-nitrobenzyl)isoindoline-1,3-dione⁴³ (500 mg, 1.77 mmol) in a mixture of tetrahydrofuran and methanol (5 mL/0.5 mL) was added sodium borohydride (114 mg, 2.65 mmol) over 10 min at -10 °C. The resulting mixture was stirred at -10 °C for 2 h. Subsequently, excess sodium borohydride was quenched with aqueous 3 N HCl (0.5 mL). Evaporation of solvents on a rotary evaporator resulted in a colorless solid which was washed with water to provide 3-hydroxy-2-(2-nitrobenzyl)isoindolin-1-one (7e) in 88% yield (443 mg): mp 98 °C; IR (KBr, cm⁻¹) 3340, 2865, 1681, 1609, 1577, 1525, 1469, 1431, 1344, 1305, 1207, 1060, 961, 925, 858, 788, 748, 728; ¹H NMR (400 MHz, DMSO- d_6 /CCl₄ 1/1) δ 8.07 (d, J = 8.0 Hz, 1H), 7.73–7.52 (m, 6H), 7.40 (d, J = 7.2 Hz, 1H), 6.69 (d, J = 8.0 Hz, 1H), 5.78 (d, J = 8.0 Hz, 1H), 5.06 (d, J = 17.3 Hz, 1H), 4.88 (d, J = 17.3 Hz, 1H; ¹³C NMR (100 MHz, DMSO- $d_6/\text{CCl}_4 1/1$) δ 166.6 (C), 147.8 (C), 144.8 (C), 133.5 (CH), 132.9 (C), 132.0 (CH), 131.0 (C), 129.2 (CH), 128.8 (CH), 128.0 (CH), 124.5 (CH), 123.6 (CH), 122.4 (CH), 81.1 (CH), 39.8 (CH₂); HRMS (ESI) calcd for $C_{15}H_{12}N_2O_4Na$ (M + Na) 307.0695, found 307.0698.

Synthesis of 2-(2-Nitrobenzyl)-3-phenylisoindolin-1-ones 10h–w. The general procedure described for synthesis of 2-benzyl-3-arylisoindolin-1-ones 10a–e was followed for the synthesis of 2-(2nitrobenzyl)-3-phenylisoindolin-1-ones 10h–w.

2-(2-Nitrobenzyl)-3-phenylisoindolin-1-one (10h):



colorless solid (109 mg, 91% yield); mp 116 °C; IR (KBr, cm⁻¹) 3062, 2927, 2872, 1694, 1611, 1527, 1468, 1400, 1344, 1305, 1142, 1094, 858, 790, 738, 702; ¹H NMR (400 MHz, CDCl₃) δ 7.95 (d, *J* = 6.9 Hz, 2H), 7.54–7.48 (m, 3H), 7.40–7.36(m, 2H), 7.30–7.28 (m, 3H), 7.19–7.17 (m, 1H), 7.01 (d, *J* = 4.8 Hz, 2H), 5.40 (d, *J* = 17.2 Hz, 1H); 5.40 (s, 1H), 4.50 (d, *J* = 17.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 169.3 (C), 148.4 (C), 146.4 (C), 136.3 (C), 133.6 (C), 132.7 (CH), 132.4 (CH), 130.8 (C), 130.0 (CH), 129.3 (CH), 129.0 (CH), 128.6 (CH), 128.3 (CH), 127.6 (CH), 125.5 (CH), 123.9 (CH), 123.4 (CH), 65.0 (CH), 41.2 (CH₂); HRMS (ESI) calcd for C₂₁H₁₆N₂O₃Na (M + Na) 367.1059, found 367.1058.

2-(2-Nitrobenzyl)-3-(4-(trifluoromethyl)phenyl)isoindolin-1-one (10i):



colorless solid (134 mg, 92% yield); mp 141 °C; IR (KBr, cm⁻¹) 3070, 2935, 2860, 1699, 1617, 1528, 1395, 1326, 1166, 1125, 1067, 854, 790, 734; ¹H NMR (400 MHz, CDCl₃/CCl₄ 1/1) δ 7.94–7.89 (m, 2H), 7.56–7.48 (m, 6H), 7.40–7.38 (m, 1H), 7.19 (d, *J* = 8.1 Hz, 2H), 7.15–7.11 (m, 1H), 5.49 (s, 1H), 5.33 (d, *J* = 16.0 Hz, 1H), 4.57 (d, *J* = 16.0 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) δ 169.1 (C), 148.7 (C), 145.8 (C), 140.9 (C), 133.67 (CH), 132.64 (CH), 132.5 (C), 131.5 (C), 131.0 (CH), 130.8 (C), 129.0 (CH), 128.6 (CH), 128.0 (CH), 126.3 (m, CH), 124.9 (CH), 124.3 (CH), 123.3 (CH), 64.6 (CH), 41.3 (CH₂); HRMS (ESI) calcd for C₂₂H₁, F₃N₂O₃Na (M + Na) 435.0932, found 435.0932.

3-(4-Methoxyphenyl)-2-(2-nitrobenzyl)isoindolin-1-one (10j):



Light yellow solid (93 mg, 77% yield); mp 115 °C; IR (KBr, cm⁻¹) 3071, 2935, 2840, 1695, 1609, 1525, 1466, 1400, 1344, 1304, 1248, 1176, 1107, 1027, 787, 731; ¹H NMR (400 MHz, CDCl₃) δ 7.97–7.93 (m, 2H), 7.54–7.48 (m, 3H), 7.40–7.36 (m, 2H), 7.18–7.16 (m, 1H), 6.92 (d, *J* = 8.3 Hz, 2H), 6.80 (d, *J* = 8.3 Hz, 2H), 5.39–5.35 (m, 2H), 4.48 (d, *J* = 16.8 Hz, 1H), 3.77 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 169.2 (C), 160.0 (C), 148.4 (C), 146.7 (C), 133.7 (CH), 132.9 (C), 132.3 (CH), 130.9 (C), 129.9 (CH), 129.0 (CH), 128.6 (CH), 128.2 (CH), 127.9 (C), 125.0 (CH), 123.9 (CH), 123.4 (CH), 114.6 (CH), 64.5 (CH), 55.4 (CH₃), 41.1 (CH₂); HRMS (ESI) calcd for C₂₂H₁₈N₂O₄ Na (M + Na) 397.1164, found 397.1168.

3-(Furan-2-yl)-2-(2-nitrobenzyl)isoindolin-1-one (10k):



colorless solid (93 mg, 79% yield); mp 140 °C; IR (KBr, cm⁻¹) 1697, 1596, 1577, 1470, 1395, 1343, 1305, 1190, 1080, 858, 786, 750, 702; ¹H NMR (400 MHz, CDCl₃/CCl₄ 1/1) δ 7.97–7.93 (m, 2H), 7.61–7.43 (m, 3H), 7.38–7.32 (m, 3H), 7.22 (s, 1H), 6.24 (s, 2H), 5.55 (s, 1H), 5.25 (d, *J* = 16.9 Hz, 1H), 4.83 (d, *J* = 16.9 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) δ 168.4 (C), 148.6 (C), 148.2 (C), 143.5 (CH), 142.8 (C), 133.3 (CH), 132.7 (C), 132.1 (CH), 131.4 (C), 129.5 (CH), 128.9 (CH), 127.9 (CH), 124.7 (CH), 123.9 (CH), 123.2 (CH), 110.5 (CH), 110.2 (CH), 58.3 (CH), 41.5 (CH₂); HRMS (ESI) calcd for C₁₉H₁₄N₂O₄Na (M + Na) 357.0851, found 357.0851.

3-(4-Fluorophenyl)-2-(2-nitrobenzyl)isoindolin-1-one (10l):



Light yellow solid (116 mg, 91% yield); mp 157 °C; IR (KBr, cm⁻¹) 3071, 2930, 1695, 1607, 1527, 1469, 1397, 1342, 1226, 1158, 1097,

856, 788, 737; ¹H NMR (400 MHz, CDCl₃) δ 7.94–7.91 (m, 2H), 7.53–7.48 (m, 3H), 7.40–7.35 (m, 2H), 7.16–7.13 (m, 1H), 7.01– 6.93 (m, 4H), 5.40 (s, 1H), 5.35 (d, *J* = 16.4 Hz, 1H), 4.50 (d, *J* = 16.4 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 169.1 (C), 162.8 (d, *J* = 241 Hz, C), 148.3 (C), 146.2 (C), 133.6 (CH), 132.5 (C), 132.4 (CH), 132.1 (d, *J* = 3 Hz, C), 130.7 (C), 130.1 (CH), 129.4 (d, *J* = 9 Hz, CH), 128.7 (CH), 128.4 (CH), 124.9 (CH), 123.9 (CH), 123.3 (CH), 116.2 (d, *J* = 22 Hz, CH), 64.2 (CH), 41.0 (CH₂); HRMS (ESI) calcd for C₂₁H₁₅FN₂O₃Na (M + Na) 385.0964, found 385.0964.

3-(3,5-Difluorophenyl)-2-(2-nitrobenzyl)isoindolin-1-one (10m):



light yellow solid (108 mg, 81% yield); mp 167 °C; IR (KBr, cm⁻¹) 1713, 1650, 1574, 1530, 1464, 1409, 1333, 1116, 855, 790, 729; ¹H NMR (400 MHz, CDCl₃) δ 7.96 (d, J = 6.8 Hz, 2H), 7.56–7.51 (m, 3H), 7.45–7.39 (m, 2H), 7.18 (d, J = 4.8 Hz, 1H), 6.74 (t, J = 8.5 Hz, 1H), 6.61 (d, J = 5.6 Hz, 2H), 5.41–5.37 (m, 2H), 4.57 (d, J = 16.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 169.2 (C), 16.3.5 (d, J = 249 Hz, C) 163.4 (d, J = 250 Hz, C), 148.5 (C), 145.3 (C), 140.8 (C), 133.8 (CH), 132.7 (CH), 132.2 (C), 130.54 (C), 130.48 (CH), 129.1 (CH), 128.7 (CH), 125.1 (CH), 124.3 (CH), 123.2 (CH), 110.4 (t, J = 18 Hz, CH), 104.5 (t, J = 25 Hz, CH), 64.2 (CH), 41.3 (CH₂); HRMS (ESI) calcd for C₂₁H₁₄F₂N₂O₃Na (M + Na) 403.0870, found 403.0871

3-(4-Chlorophenyl)-2-(2-nitrobenzyl)isoindolin-1-one (10n):



colorless solid (109 mg, 81% yield); mp 151 °C; IR (KBr, cm⁻¹) 1698, 1612, 1526, 1490, 1468, 1395, 1345, 1307, 1088, 789, 748, 719; ¹H NMR (400 MHz, CDCl₃/CCl₄ 1/1) δ 7.92 (d, *J* = 7.2 Hz, 2H), 7.52–7.39 (m, 5H), 7.27–7.24 (m, 2H), 7.14–7.12 (m, 1H), 6.98 (d, *J* = 8.4 Hz, 2H), 5.39 (s, 1H), 5.33 (d, *J* = 16.4 Hz, 1H), 4.51 (d, *J* = 16.4 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) δ 169.0 (C), 148.5 (C), 146.0 (C), 135.0 (C), 134.9 (C), 133.5 (C), 132.5 (CH), 132.4 (CH), 120.8 (CH), 124.1 (CH), 123.3 (CH), 64.3 (CH), 41.1 (CH₂); HRMS (ESI) calcd for C₂₁H₁₅ClN₂O₃Na (M + Na) 401.0669, found 401.0673.

3-(3-Chlorophenyl)-2-(2-nitrobenzyl)isoindolin-1-one (10o):



colorless solid (118 mg, 88% yield); mp 165 °C; IR (KBr, cm⁻¹) 3063, 2918, 2859, 1697, 1526, 1470, 1396, 1344, 1305, 1191, 787, 703; ¹H NMR (400 MHz, CDCl₃/CCl₄ 1/1) δ 7.94–7.91 (m, 2H), 7.53–7.49 (m, 3H), 7.44–7.38 (m, 2H), 7.25–7.15 (m, 3H), 6.98–6.96 (m, 2H), 5.39 (s, 1H), 5.35 (d, *J* = 16.8 Hz, 1H),(d, *J* = 16.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) δ 169.1 (C), 148.5 (C), 145.8 (C), 138.6 (C), 135.2 (C), 133.6 (CH), 132.5 (CH), 132.4 (C), 130.7 (C), 130.5 (CH), 125.8 (CH), 124.9 (CH), 124.1 (CH), 123.3 (CH), 64.5 (CH), 41.2 (CH₂); HRMS (ESI) calcd for C₂₁H₁₅ClN₂O₃Na (M + Na) 401.0669, found 401.0664.

3-(4-Bromophenyl)-2-(2-nitrobenzyl)isoindolin-1-one (10p):



colorless solid (126 mg, 85% yield); mp 114 °C; IR (KBr, cm⁻¹) 1697, 1613, 1526, 1469, 1396, 1344, 1306, 1072, 1011, 788, 745; ¹H NMR (400 MHz, CDCl₃) δ 7.93 (d, J = 8.1 Hz, 2H), 7.54–7.49 (m, 3H), 7.42–7.37 (m, 4H), 7.15–7.13 (m, 1H), 6.91 (d, J = 7.0 Hz, 2H), 5.38–5.34 (m, 2H), 4.51 (d, J = 16.8 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 169.2 (C), 148.5 (C), 145.9 (C), 135.5 (C), 133.7 (CH), 132.5 (CH), 132.4 (2C, C,CH), 130.7 (C), 130.3 (CH), 129.3 (CH), 128.8 (CH), 128.5 (CH), 125.0 (CH), 124.1 (CH), 123.3 (CH), 123.0 (C), 64.4 (CH), 41.2 (CH₂); HRMS (ESI) calcd for C₂₁H₁₅BrN₂O₃Na (M + Na) 445.0164, found 445.0162.

2-(2-Nitrobenzyl)-3-(p-tolyl)isoindolin-1-one (10q):



colorless solid (113 mg, 89% yield); mp 135 °C; IR (KBr, cm⁻¹) 1696, 1609, 1527, 1468, 1400, 1345, 1305, 1200, 1158, 1095, 858, 787, 734, 706; ¹H NMR (400 MHz, CDCl₃/CCl₄ 1/1) δ 7.92 (d, *J* = 6.9 Hz, 2H), 7.59–7.30 (m, 5H), 7.20–7.03 (m, 3H), 6.89 (d, *J* = 7.7 Hz, 2H), 5.38–5.33 (m, 2H), 4.45 (d, *J* = 16.8 Hz, 1H), 2.31 (s, 3H); ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) δ 169.0 (C), 148.5 (C), 146.6 (C), 138.7 (C), 133.5 (CH), 133.2 (C), 132.9 (C), 132.2 (CH), 131.0 (C), 130.1 (CH), 129.9 (CH), 128.5 (CH), 128.1 (CH), 127.6 (CH), 124.9 (CH), 123.9 (CH), 123.4 (CH), 64.7 (CH), 41.0 (CH₂), 21.3 (CH₃); HRMS (ESI) calcd for C₂₂H₁₈N₂O₃Na (M + Na) 381.1215, found 381.1215.

2-(2-Nitrobenzyl)-3-(m-tolyl)isoindolin-1-one (10r):



colorless solid (109 mg, 86% yield); mp 129 °C; IR (KBr, cm⁻¹), 3049, 2922, 1694, 1609, 1578, 1526, 1469, 1397, 1342, 1304, 1200, 1095, 857, 787, 770, 730, 706; ¹H NMR (400 MHz, CDCl₃/CCl₄ 1/1) δ 7.93–7.91 (m, 2H), 7.53–7.47 (m, 3H), 7.40–7.35 (m, 2H), 7.18–7.13 (m, 2H), 7.08–7.06 (m, 1H), 6.81 (d, *J* = 7.6 Hz, 1H), 6.77 (s, 1H), 5.38 (d, *J* = 16.8 Hz, 1H), 5.35 (s, 1H), 5.49 (d, *J* = 16.8 Hz, 1H), 2.25 (s, 3H); ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) δ 169.2 (C), 148.4 (C), 146.4 (C), 139.0 (C), 136.1 (C), 133.5 (CH), 132.8 (C), 132.3 (CH), 130.9 (C), 130.0 (CH), 129.7 (CH), 129.1 (CH), 128.5 (CH), 128.2 (CH), 128.0 (CH), 124.9 (CH), 124.8 (CH), 123.9 (CH), 123.4 (CH), 65.0 (CH), 41.1 (CH₂), 21.4 (CH₃); HRMS (ESI) calcd for C₂₂H₁₈N₂O₃Na (M + Na) 381.1215, found 381.1214. *3*-(2,3-Dimethoxyphenyl)-2-(2-nitrobenzyl)isoindolin-1-one (**105**):



light yellow solid (113 mg, 79% yield); mp 122 °C; IR (KBr, cm⁻¹) 3066, 2937, 2837, 1697, 1605, 1522, 1465, 1399, 1263, 1141, 1027, 858, 788, 730; ¹H NMR (400 MHz, $CDCl_3/CCl_4$ 1/1) δ 7.84–7.80

(m, 2H), 7.42–7.36 (m, 4H), 7.30–7.26 (m, 1H), 7.10–7.08 (m, 1H), 6.68 (d, J = 8.0 Hz, 1H), 6.56 (d, J = 8.0 Hz, 1H), 6.27 (s, 1H), 5.25–5.21 (m, 2H), 4.47 (d, J = 17.6 Hz, 1H), 3.75 (s, 3H), 3.61 (s, 3H); ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) δ 168.7 (C), 149.6 (C), 149.5 (C), 148.5 (C), 146.4 (C), 133.2 (CH), 132.7 (C), 132.1 (CH), 130.8 (C), 130.2 (CH), 128.4 (CH), 128.3 (C), 128.0 (CH), 124.6 (CH), 123.7 (CH), 123.2 (CH), 120.5 (CH), 111.4 (CH), 109.9 (CH), 64.8 (CH), 55.75 (CH₃), 55.71 (CH₃), 40.8 (CH₂); HRMS (ESI) calcd for C₂₃H₂₀N₂O₅Na (M + Na) 427.1270, found 427.1269.

3-(2,5-Dimethoxyphenyl)-2-(2-nitrobenzyl)isoindolin-1-one (10t):



colorless solid (116 mg, 81% yield); mp 130 °C; IR (KBr, cm⁻¹) 3002, 2936, 2836, 1694, 1526, 1468, 1340, 1280, 1218, 1047, 856, 789, 748; ¹H NMR (400 MHz, CDCl₃/CCl₄ 1/1) δ 7.93 (d, *J* = 6.8 Hz, 2H), 7.50–7.46 (m, 3H), 7.36–7.28 (m, 3H), 6.78–6.72 (m, 2H), 6.17 (s, 1H), 6.03 (s, 1H), 5.38 (d, *J* = 16.8 Hz, 1H), 5.57 (d, *J* = 16.8 Hz, 1H), 3.70 (s, 3H), 3.58 (s, 3H); ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) δ 169.3 (C), 153.9 (C), 151.9 (C), 148.4 (C), 146.4 (C), 133.4 (CH), 133.3 (C), 132.1 (CH), 131.4 (C), 129.7 (CH), 128.3 (CH), 127.9 (CH), 125.3 (C), 124.7 (CH), 123.9 (CH), 123.4 (CH), 114.4 (CH), 112.8 (CH), 111.9 (CH), 57.4 (CH), 55.8 (CH₃), 55.5 (CH₃), 41.2 (CH₂); HRMS (ESI) calcd for C₂₃H₂₀N₂O₃Na (M + Na) 427.1270, found 427.1273.

3-(2,6-Dimethoxyphenyl)-2-(2-nitrobenzyl)isoindolin-1-one (10u):



colorless solid (93 mg, 74% yield); mp 154 °C; IR (KBr, cm⁻¹) 2933, 2831, 1693, 1528, 1462, 1392, 1284, 1217, 1032, 853, 783, 747; ¹H NMR (400 MHz, CDCl₃) δ 7.98–7.94 (m, 2H), 7.51–7.47 (m, 3H), 7.41–7.22 (m, 3H), 6.53 (s, 1H), 6.47–6.26 (m, 2H), 5.98 (s, 1H), 5.38 (d, *J* = 17.0 Hz, 1H), 4.54 (d, *J* = 17.1 Hz, 1H), 3.76 (s, 3H), 3.68 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 169.4 (C), 161.1 (C), 158.9 (C), 148.3 (C), 146.8 (C), 133.5 (C), 133.5 (CH), 132.1 (CH), 131.5 (C), 129.3 (CH), 128.2 (CH), 127.8 (CH), 124.7 (CH), 123.7 (CH), 123.4 (CH), 116.4 (C), 105.3 (CH), 98.4 (2 × CH), 57.1 (CH), 55.4 (CH₃), 55.3 (CH₃), 41.2 (CH₂); HRMS (ESI) calcd for C₂₃H₂₀N₂O₅Na (M + Na) 427.1270, found 427.1275.

(E)-2-(2-Nitrobenzyl)-3-styrylisoindolin-1-one (10v):



viscous solid (82 mg, 63% yield); IR (KBr, cm⁻¹) 3058, 3028, 2960, 2927, 2869, 1696, 1609, 1527, 1469, 1402, 1345, 1305, 1149, 1084, 970, 857, 751, 698; ¹H NMR (400 MHz, CDCl₃/CCl₄ 1/1) δ 7.95 (dd, *J* = 8.2, 1.2 Hz, 2H), 7.90 (dd, *J* = 6.7, 1.0 Hz, 2H), 7.59–7.49 (m, 6H), 7.44 (d, *J* = 6.8 Hz, 2H), 7.37 (t, *J* = 7.5 Hz, 4H), 7.28–7.26 (m, 2H), 7.26–7.23 (m, 3H), 6.75 (d, *J* = 15.7 Hz, 1H), 5.71 (dd, *J* = 15.7, 9.2 Hz, 1H), 5.31 (d, *J* = 16.8 Hz, 1H), 4.99 (d, *J* = 9.2 Hz, 1H), 4.91 (d, *J* = 16.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) δ 168.7 (C), 148.7 (C), 144.8 (C), 136.6 (C), 135.6 (CH), 133.6 (CH), 133.3 (C), 132.2 (CH), 131.5 (C), 130.2 (CH), 129.0 (CH), 128.8 (CH), 128.7 (CH), 128.3 (CH), 126.9 (CH), 125.1 (CH), 125.0 (CH),

124.2 (CH), 123.5 (CH), 64.2 (CH), 41.3 (CH₂); HRMS (ESI) calcd for $C_{23}H_{18}N_2O_3Na$ (M + Na) 393.1215, found 393.1215.

(E)-3-(Hex-1-en-1-yl)-2-(2-nitrobenzyl)isoindolin-1-one (10w):



viscous solid (103 mg 84%); IR (KBr, cm⁻¹) 2958, 2928, 2864, 1767, 1696, 1613, 1467, 1401, 1343, 1304, 1223, 1148, 1099, 1049, 974, 857, 789, 731; ¹H NMR (400 MHz, CDCl₃) δ 8.01 (d, J = 8.1 Hz, 1H), 7.89 (d, J = 7.4 Hz, 1H), 7.59–7.47 (m, 3H), 7.43–7.32 (m, 3H), 5.95–5.82 (m, 1H), 5.28 (d, J = 17.0 Hz, 1H), 5.05–4.99 (m, 1H), 4.90 (d, J = 17.0 Hz, 1H), 4.79 (d, J = 9.2 Hz, 1H), 2.06–2.00 (m, 2H), 1.30–1.25 (m, 4H), 0.86 (t, J = 6.9 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.8 (C), 148.5 (C), 145.2 (C), 139.2 (CH), 133.6 (CH), 133.3 (C), 132.1 (CH), 131.3 (C), 129.7 (CH), 128.6 (CH), 128.1 (CH), 125.6 (CH), 124.9 (CH), 123.9 (CH), 123.3 (CH₃); HRMS (ESI) calcd for C₂₁H₂₂N₂O₃Na (M + Na) 373.1528, found 373.1528.

General Procedure for Deprotection of the 2-Nitrobenzyl Group for Synthesis of C(3)-Substituted Isoindolin-1-ones 11a-d: Synthesis of 3-Phenylisoindolin-1-one (11a).



A stirred solution of 2-(2-nitrobenzyl)-3-phenylisoindolin-1-one (**10h**; 50 mg, mmol) in CH₃CN/H₂O (1/1; 10 mL) in a Pyrex test tube of 20 mL capacity was exposed to light irradiation with emission maximum at 370 nm emitted by UV-LED ($4 \times 3 \mu$ W) lamps (see the Supporting Information for an image of the reactor built in-house). After completion of deprotection (4 h) the solvent was removed under reduced pressure, and the resulting faintly white solid was subjected to column chromatography (silica gel, gradient elution with an increasing volume of EtOAc in hexanes) to separate 3-phenylisoindolin-1-one (**11a**) from nitrosobenzaldehyde. The isoindolin-1-one **11a** was obtained as a colorless solid (28 mg, 93% yield): mp 219 °C (reported 219 °C). Spectral data (IR, ¹H NMR and ¹³C NMR, and DEPT) of **11a** matched those reported by Slavov and coworkers.⁴⁴

3-(4-Methoxyphenyl)isoindolin-1-one (11b):



colorless solid (28 mg, 88% yield); mp 155 °C; IR (KBr, cm⁻¹) 3200, 3074, 2955, 1732, 1697, 1589, 1514, 1346, 1246, 732; ¹H NMR (400 MHz, CDCl₃/CCl₄ 1/1) δ 7.92–7.86 (m, 1H), 7.53–7.43 (m, 2H), 7.21 (dd, *J* = 7.5, 0.9 Hz, 1H), 7.16 (d, *J* = 8.7 Hz, 2H), 6.87 (d, *J* = 8.8 Hz, 2H), 6.63 (s, 1H), 5.58 (s, 1H), 3.79 (s, 3H). ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) δ 171.0 (C), 159.9 (C), 148.3 (C), 132.4 (CH), 130.9 (C), 130.3 (C), 128.4 (CH), 128.2 (CH), 123.9 (CH), 123.4 (CH), 114.5 (CH), 60.4 (CH₃), 55.4 (CH); HRMS (ESI) calcd for C₁₄H₁₁FNO (M + H) 240.1019, found 240.1019.

3-(4-Fluorophenyl)isoindolin-1-one (11c):



colorless solid (27 mg, 87% yield); mp 179 °C; IR (KBr, cm⁻¹) 3194, 3068, 2837, 1685, 1610, 1512, 1465, 1358, 1247, 1178, 1033, 731; ¹H NMR (400 MHz, CDCl₃/CCl₄ 1/1) 7.83 (m, 1H), 7.63 (bs, 1H), 7.50–7.49 (m, 2H), 7.25–7.18 (m, 3H), 7.01 (t, J = 8.8 Hz, 2H), 5.60 (s, 1H); ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) 171.3 (C), 162.9 (d, J = 247 Hz, C), 148.0 (C), 134.4 (d, J = 3 Hz, C), 132.4 (CH), 131.1 (C), 128.7 (CH), 128.6 (d, J = 6 Hz, CH), 124.1 (CH), 123.3 (CH), 116.2 (d, J = 21 Hz, CH), 60.4 (CH); HRMS (ESI) calcd for C₁₄H₁₁FNO (M + H) 228.0819, found 228.0813.

3-(3-Chlorophenyl)isoindolin-1-one (11d):



colorless solid (28 mg, 88% yield); mp 157 °C; IR (KBr, cm⁻¹) 3191, 3071, 2924, 1690, 1552, 1469, 1352, 1191, 1139, 1088, 784, 749, 705; ¹H NMR (400 MHz, CDCl₃/CCl₄ 1/1) 7.51–7.48 (m, 1H), 7.29–7.27 (m, 2H), 7.25–7.15 (m, 6H), 6.97 (bs, 1H), 5.58 (s, 1H); ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) 171.6 (C), 147.5 (C), 140.7 (C), 135.2 (C), 132.6 (CH), 131.0 (C), 130.5 (CH), 128.9 (CH), 128.8 (CH), 127.1 (CH), 125.0 (CH), 124.2 (CH), 123.3 (CH), 60.5 (CH); HRMS (ESI) calcd for $C_{14}H_{11}$ CINO (M + H) 244.0529, found 244.0524.

Preparation of 2-(2-Bromo-4,5-dimethoxyphenethyl)isoindoline-1,3-dione (16).



A suspension of phthalic anhydride (600 mg, 4 mmol) and 2-(2bromo-4,5-dimethoxyphenyl)ethan-1-amine (1.036 g, 4 mmol) in 15 mL of toluene in an oven-dried 25 mL round-bottom flask fitted with a Dean-Stark apparatus was heated to reflux until a clear solution of the product was obtained (6 h). The condensation worked better in toluene reflux with a Dean-Stark apparatus in comparison to the condensation with phthaloyl dichloride in acetonitrile in the presence of Hunig's base as described by DiMagno and co-workers.³⁹ After completion of the reaction the mixture was concentrated under reduced pressure to give a residue which was subjected to column chromatography (silica gel, gradient elution with increasing volume of EtOAc in hexanes) to furnish 16 as a colorless solid (1.31 g, 82% yield): mp = 143 °C; IR (KBr, cm⁻¹) 1713, 1583, 1467, 1386, 1121, 1084, 883, 768, 712; ¹H NMR (400 MHz, CDCl₃) δ 7.81 (dd, J = 5.5, 3.0 Hz, 2H), 7.69 (dd, J = 5.4, 3.1 Hz, 2H), 6.98 (s, 1H), 6.66 (s, 1H), 3.94 (t, J = 8.0 Hz, 2H), 3.82 (s, 3H), 3.70 (s, 3H), 3.06 (t, J = 8 0 Hz, 2H); ¹³C NMR $(100 \text{ MHz}, \text{CDCl}_3) \delta 168.2 \text{ (C)}, 148.48 \text{ (C)}, 148.44 \text{ (C)}, 134.0 \text{ (CH)},$ 132.1(C), 129.4 (C), 123.3 (CH), 115.69 (CH), 114.5 (C), 113.3 (CH), 56.1 (CH₃), 56.0 (CH₃), 37.7 (CH₂), 34.4 (CH₂).

2-(4,5-Dimethoxy-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)phenethyl)isoindoline-1,3-dione (17).



A viscous solution of 2-(2-bromo-4,5-dimethoxyphenethyl)isoindoline-1,3-dione (16; 300 mg, 0.75 mmol), bis(pinacolato)diboron (222 mg, 0.90 mmol), NaOAc (242 mg, 3 mmol), and Pd(PPh₃)₂Cl₂ (26 mg, 0.08 mmol) in PEG 400 (3 mL) was stirred in an oil bath at 80 °C for 3 h under an atmosphere of nitrogen. After completion of the reaction (TLC, 20% EtOAc in hexanes, $R_f = 0.5$), the cooled mixture was extracted with methyl *tert*-butyl ether (MTBE, 2×10 mL). The ether layer was washed with brine and dried over anhydrous sodium sulfate. The crude product obtained after removal of MTBE was purified by column chromatography (silica gel, gradient elution with increasing volume of EtOAc in hexanes), resulting in 17 as a colorless solid (269 mg, 84% yield): mp 124 °C; IR (KBr, cm⁻¹) 3005, 2941, 1712, 1500, 1394, 1361, 1255, 1213, 1163, 1097, 999, 869, 800, 717; ¹H NMR (400 MHz, $CDCl_3$) δ 7.79 (dd, J = 5.4, 3.0 Hz, 2H), 7.67 (dd, J = 5.4, 3.0 Hz, 2H), 7.28 (s, 1H), 6.65 (s, 1H), 4.00-3.92 (m, 2H),3.89 (s, 3H), 3.73 (s, 3H), 3.30–3.22 (m, 2H), 1.37 (s, 12H); ^{13}C NMR (100 MHz, CDCl₃) δ 168.3 (C), 150.9 (C), 146.8 (C), 139.1 (C), 133.8 (CH), 132.2 (C), 123.1 (CH), 118.3 (CH), 113.1 (CH), 83.6 (C), 55.9 (CH₃), 55.7 (CH₃), 40.1 (CH₂), 33.9 (CH₂), 24.9 (CH₃); HRMS (ESI) calcd for C₂₄H₂₈NO₆BNa (M + Na) 460.1908, found 460.1906.

2-(4,5-Dimethoxy-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)phenethyl)-3-hydroxyisoindolin-1-one (18).



To a stirred solution of 2-(4,5-dimethoxy-2-(4,4,5,5-tetramethyl-1,3,2dioxaborolan-2-yl)phenethyl)isoindoline-1,3-dione (17; 200 mg, 0.46 mmol) in a mixture of tetrahydrofuran (3 mL) and methanol (0.3 mL) kept in an ice-water bath was added sodium borohydride (34 mg, 0.92 mmol) in three portions over 10 min. The reaction mixture was then stirred at 0-5 °C for 20 min, by which time the reduction was complete (TLC). Excess sodium borohydride was quenched with water (1 mL). Removal of THF and MeOH under reduced pressure in a rotary evaporator resulted in a suspension of a white solid in residual water. Filtration of the solid followed by washing with water (5 mL) provided 18 as a colorless solid in 96% yield (192 mg): mp 138 °C; IR (KBr, cm⁻¹) 3171, 2943, 1660, 1589, 1516, 1440, 1263, 1143, 1024, 819, 744, 698; ¹H NMR (400 MHz, CDCl₃/CCl₄ 1/1) δ 7.57-7.43 (m, 3H), 7.36-7.26 (m, 1H), 7.25 (d, J = 1.1 Hz, 1H), 6.64 (d, J = 1.8 Hz, 1H), 5.56 (d, J = 2.3 Hz, 1H), 3.87 (d, J = 1.0 Hz, 3H), 3.73 (d, J = 1.8 Hz, 3H), 3.65-3.56 (m, 1H), 3.53-3.43 (m, 1H), 3.21-3.11 (m, 1H), 3.06-2.96 (m, 1H), 1.33 (s, 12H); ¹³C NMR (100 MHz, $CDCl_3/CCl_4$ 1/1) δ 167.3 (C), 151.4 (C), 146.9 (C), 144.0 (C), 139.9 (C), 132.0 (C), 131.9 (CH), 129.5 (CH), 123.3 (CH), 123.2 (CH), 118.8 (CH), 113.1 (CH), 83.7 (C), 82.1 (CH), 55.9 (CH₃), 55.7 (CH₃), 42.0 (CH₂), 34.2 (CH₂), 24.9 (CH_3) , 24.8 (CH_3) ; HRMS (ESI) calcd for $C_{24}H_{30}NO_6BNa$ (M + Na) 462.2064, found 462.2064.

2,3-Dimethoxy-5,12b-dihydroisoindolo[1,2-*a*]isoquinolin-8(6*H*)-one (19).



By following the general procdure described for the copper-mediated coupling reactions described earlier, intramolecular coupling in 2-(4,5-dimethoxy-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)phenethyl)-3-hydroxyisoindolin-1-one (**18**; 100 mg, 0.23 mmol) in the presence of $Cu(OTf)_2$ (8 mg, 0.023 mmol) and Na_2CO_3 (12 mg, 0.11 mmol) in DCE (2 mL) furnished 2,3-dimethoxy-5,12b-dihydroisoindolo[1,

2-*a*]isoquinolin-8(6*H*)-one (**19**) as a colorless solid in 89% yield (59 mg): mp 60 °C; IR (KBr, cm⁻¹) 2912, 1678, 1518, 1417, 1253, 1228, 1205, 1093, 724; ¹H NMR (400 MHz, CDCl₃/CCl₄ 1/1) δ 7.85 (d, *J* = 7.5 Hz, 1H), 7.79 (dd, *J* = 7.7, 0.6 Hz, 1H), 7.58 (td, *J* = 7.5, 1.2 Hz, 1H), 7.47 (t, *J* = 7.5 Hz, 1H), 7.08 (s, 1H), 6.62 (s, 1H), 5.58 (s, 1H), 4.51–4.45 (m, 1H), 3.91 (s, 3H), 3.83 (s, 3H), 3.41–3.38 (m, 1H), 3.02–2.94 (m, 1H), 2.74 (dt, *J* = 15.8, 4.0 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃/CCl₄ 1/1) δ 167.9 (C), 148.6 (C), 148.1 (C), 144.8 (C), 132.9 (C), 131.6 (CH), 128.6 (CH), 127.1 (C), 126.2 (C), 124.2 (CH), 123.1 (CH), 112.2 (CH), 109.0 (CH), 59.1 (CH), 56.3 (CH₃), 56.0 (CH₃), 38.3 (CH₂), 29.2 (CH₂); HRMS (ESI) calcd for C₁₈H₁₈NO₃ (M + H) 296.1287, found 296.1276.

ASSOCIATED CONTENT

S Supporting Information

Figures and a CIF file giving ¹H and ¹³C NMR and DEPT-135 spectra for all compounds prepared, an ORTEP plot of the X-ray structure of **10h**, a picture of the home-built UV reactor, and crystal data for **10h**. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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REFERENCES

 (1) (a) Speck, K.; Magauer, T. Beilstein J. Org. Chem. 2013, 9, 2048–2078.
 (b) Heugebaert, T. S. A.; Roman, B. I.; Stevens, C. V. Chem. Soc. Rev. 2012, 41, 5626–5640.
 (c) Subbarayappa, A.; Patoliya, P. U. Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem. 2009, 48, 545–552.
 (d) Kundu, N. G.; Khan, M. W.; Mukhopadhyay, R. J. Indian Chem. Soc. 2001, 78, 671–688.

(2) (a) Meng, Z.-H.; Liao, L.-H.; Pommier, Y. Curr. Top. Med. Chem.
2003, 3, 305–320. (b) Garcia-Carbonero, R.; Supko, J. G. Clin. Cancer Res. 2002, 8, 641–661. (c) Pendrak, I.; Barney, S.; Wittrock, R.; Lambert, D. M.; Kingsbury, W. D. J. Org. Chem. 1994, 59, 2623–2625.
(d) Sundberg, R. J. In Comprehensive Heterocyclic Chemistry. Pyrroles and Their Benzo Derivatives: (iii) Synthesis and Application; Bird, C. W., Cheeseman, G. W. H., Eds.; Elsevier: Amsterdam, 1984; Vol 4, pp 313–376.

(3) Bentley, K. W. In *The Isoquinoline Alkaloids*; Ravindranath, B., Ed.; Harwood Academic: Amsterdam, 1998; pp 361–375.

(4) Tejesvi, M. V.; Pirttilä, A. M. In *Endophytes of Forest Trees: Biology* and Applications. Forestry Sciences; Pirttilä, A. M., Frank, A. C., Eds.; Springer: Berlin, 2011; Vol. 80, pp 302.

(5) Chen, Z.-L.; Zhu, D.-Y. In *The Alkaloids: Chemistry and Pharmacology*; Brossi, A., Ed.; Academic Press: New York, 1987, Vol. 31, pp 29–62.

(6) (a) Oak, J. N.; Oldenhof, J.; Van Tol, H. H. M. *Eur. J. Pharmacol.* 2000, 405, 303–327. (b) Belliotti, T. R.; Brink, W. A.; Kesten, S. R.; Rubin, J. R.; Wustrow, D. J.; Zoski, K. T.; Whetzel, S. Z.; Corbin, A. E.; Pugsley, T. A.; Heffner, T. G.; Wise, L. D. *Bioorg. Med. Chem. Lett.* 1998, 8, 1499–1502.

(7) (a) Zhuang, Z.; Kung, M.; Mu, M.; Kung, H. F. J. Med. Chem. 1998, 41, 157–166. (b) Norman, M. H.; Minick, D. J.; Rigdon, G. C. J. Med. Chem. 1996, 39, 149–157.

(8) Ferland, J.-M.; Demerson, C. A.; Humber, L. G. Can. J. Chem. 1985, 63, 361-365.

(9) Lippmann, W. U.S. Patent 4,267,189, 1981. Chem. Abstr. 1981, 95, 61988m.

(10) (a) Wada, T.; Fukuda, N. Pharmacol., Biochem. Behav. **1992**, 41, 573–579. (b) Wada, T.; Fukuda, N. Psychopharmacology **1991**, 103, 314–322.

(11) (a) Hyster, T. K.; Ruhl, K. E.; Rovis, T. J. Am. Chem. Soc. 2013, 135, 5364–5367. (b) Fujioka, M.; Morimoto, T.; Tsumagari, T.; Tanimoto, H.; Nishiyama, Y.; Kakiuchi, K. J. Org. Chem. 2012, 77, 2911–2923. (c) Augner, D.; Gerbino, D. C.; Slavov, N. Org. Lett. 2011, 13, 1629–1631. (d) Shacklady-McAtee, D. M.; Dasgupta, S.; Watson, M. P. Org. Lett. 2011, 13, 3490–3493. (e) Slavov, N.; Cvengroš, J.; Neudörfl, J.-M.; Schmalz, H.-G. Angew. Chem., Int. Ed. 2010, 49, 7588–7591.

(12) Zhang, Y.; DeSchepper, D. J.; Gilbert, T. M.; Sai, K. K. S.; Klumpp, D. A. Chem. Commun. 2007, 4032–4034.

(13) Klumpp, D. A.; Zhang, Y.; Connor, M. J. O.; Esteves, P. M.; De Almeida, L. S. Org. Lett. **200**7, *9*, 3085–3088.

(14) Allin, S. M.; Northfield, C. J.; Page, M. I.; Slawin, A. M. Z. *Tetrahedron Lett.* **1998**, *39*, 4905–4908.

(15) Hamersma, J. A. M.; Speckamp, W. N. Tetrahedron 1982, 38, 3255-3266.

(16) Pin, F.; Comesse, S.; Garrigues, B.; Marchalín, S.; Daïch, A. J. Org. Chem. 2007, 72, 1181–1191.

(17) Ben Othman, R.; Affani, R.; Tranchant, M.-J.; Antoniotti, S.; Dalla, V.; Duñach, E. Angew. Chem., Int. Ed. 2010, 49, 776–780.

(18) (a) Maity, A. K.; Roy, S. Adv. Synth. Catal. 2014, 356, 2627–2642. (b) Maity, A. K.; Roy, S. J. Org. Chem. 2012, 77, 2935–41.

(19) Boiaryna, L.; El Mkaddem, M. K.; Taillier, C.; Dalla, V.; Othman, M. Chem. Eur. J. 2012, 18, 14192–14200.

(20) (a) Miyaura, N. In Metal-Catalyzed Cross-Coupling Reactions; Meijere, A. D., Diederich, F., Eds.; Wiley: Weinheim, Germany, 2004; pp 41–123. Reviews: (b) Magano, J.; Dunetz, J. R. Chem. Rev. 2011, 111, 2177–2250. (c) McGlacken, G. P.; Bateman, L. M. Chem. Soc. Rev. 2009, 38, 2447–2464. (d) Torborg, C.; Beller, M. Adv. Synth. Catal. 2009, 351, 3027–3043. (e) Darses, S.; Genet, J.-P. Chem. Rev. 2008, 108, 288–325. (f) Alberico, D.; Scott, M. E.; Lautens, M. Chem. Rev. 2007, 107, 174–238. (g) Bedford, R. B.; Cazin, C. S. J.; Holder, D. Coord. Chem. Rev. 2004, 248, 2283–2321. (h) Bellina, F.; Carpita, A.; Rossi, R. Synthesis 2004, 2004, 2419–2440. (i) Hassan, J.; Se, M.; Gozzi, C.; Schulz, E.; Lemaire, M. Chem. Rev. 2002, 102, 1359–1469. (j) Miyaura, N.; Suzuki, A. Chem. Rev. 1995, 95, 2457–2483.

(21) (a) Zhou, Y.; You, W.; Smith, K. B.; Brown, M. K. Angew. Chem. 2014, 126, 3543–3547. (b) Chen, L.; Lang, H.; Fang, L.; Zhu, M.; Liu, J.; Yu, J.; Wang, L. Eur. J. Org. Chem. 2014, 2014, 4953–4957. (c) Ke, H.; Chen, X.; Zou, G. J. Org. Chem. 2014, 79, 7132–7140. (d) Han, F.-S. Chem. Soc. Rev. 2013, 42, 5270–5298.

(22) (a) Yamamoto,Y. In Copper-Mediated Cross-Coupling Reactions; Evano, G., Blanchard, N., Eds.; Wiley: Hoboken, NJ, 2013; pp 335– 399. (b) Qiao, J. X.; Lam, P. Y. S. Synthesis 2011, 829–856. (c) Evano, G.; Blanchard, N.; Toumi, M. Chem. Rev. 2008, 108, 3054–3131.
(d) Ley, S. V.; Thomas, A. W. Angew. Chem., Int. Ed. 2003, 42, 5400– 5449. (e) Lipshutz, B. H.; Sengupta, S. Org. React. 1992, 41, 135.
(f) Erdik, E. Tetrahedron 1984, 40, 641. (g) Posner, G. H. Org. React. 1975, 22, 253.

(23) (a) Flores-Rizo, J. O.; Esnal, I.; Osorio-Martínez, C. A.; Gómez-Durán, C. F. A.; Bañuelos, J.; López Arbeloa, I.; Pannell, K. H.; Metta-Magaña, A. J.; Peña-Cabrera. *Eur. J. Org. Chem.* 2013, 78, 5867–5877.
(b) Qiao, J. X.; Lam, P. Y. S. In *Boronic Acids: Recent Advances in the Chan-Lam Coupling Reaction*, 2nd ed.; Hall, D. G., Ed.; Wiley-VCH: Weinheim, Germany, 2011; pp 315–361. (c) Niu, J.; Zhou, H.; Li, Z.; Xu, J. J. Org. Chem. 2008, 73, 7814–7817.

(24) Ullmann, F.; Bielecki, J. Chem. Ber. 1901, 34, 2174-2185.

(25) (a) Yang, C.-T.; Zhang, Z.-Q.; Liu, Y.-C.; Liu, L. Angew. Chem, Int. Ed. **2011**, 50, 3904–3907. (b) Li, J.-H.; Li, J.-L.; Wang, D.-P.; Pi, S.-F.; Xie, Y.-X.; Zhang, M.-B.; Hu, X.-C. J. Org. Chem. **2007**, 72, 2053–2057. (c) Li, J.-H.; Wang, D.-P. Eur. J. Org. Chem. **2006**, 2063– 2066. (d) Thathagar, M. B.; Beckers, J.; Rothenberg, G. J. Am. Chem. Soc. 2002, 124, 11858-11859.

(26) Hall, D. G. In Boronic Acids: Structure, Properties, and Preparation of Boronic Acid Derivatives, 2nd ed.; Hall, D. G., Ed.; Wiley: Weinheim, Germany, 2011; pp 1–133.

(27) Rao, H. S. P.; Rao, A. V. B. *Eur. J. Org. Chem.* 2014, 3646–3655.
(28) (a) Harris, M. R.; Hanna, L. E.; Greene, M. G.; Moore, C. E.; Jarvo, E. R. *J. Am. Chem. Soc.* 2013, 135, 3303–3306. (b) Zhou, Q.; Srinivas, H. D.; Dasgupta, S.; Watson, M. P. *J. Am. Chem. Soc.* 2013, 135, 3307–3310. (c) Yu, J-Yi.; Kuwano, R. *Org. Lett.* 2008, 10, 973–976.

(29) Kobayashi, S.; Sugiura, M.; Kitagawa, H.; Lam, W. W.-L. Chem. Rev. 2002, 102, 2227–2302.

(30) (a) Lennox, A. J. J.; Lloyd-Jones, G. C. *Chem. Soc. Rev.* 2014, 43, 412–443. (b) Zhang, N.; Ho, D. J.; Gutsche, N.; Gupta, J.; Percec, V. J. Org. Chem. 2012, 77, 5956–5964.

(31) Billingsley, K.; Buchwald, S. L. J. Am. Chem. Soc. 2007, 129, 3358-3366.

(32) (a) Wright, J. A.; Yu, J.; Spencer, J. B. *Tetrahedron Lett.* 2001, 42, 4033–4036. (b) Cappa, A.; Marcantoni, E.; Bartoli, G.; Bellucci, M. C.; Bosco, M.; Sambri, L. J. Org. Chem. 1999, 64, 5696–5699.

(33) (a) Lemaire-Audoire, S.; Savignac, M.; Pierre, J.; Paris, C.; Bernard, J. *Tetrahedron Lett.* **1995**, *36*, 1267–1270. (b) Mereyala, H. B.; Guntha, S. *Tetrahedron Lett.* **1993**, *34*, 6929–6930.

(34) (a) Givens, R. S.; Conrad, P. G., II; Yousef, A. L.; Lee, J.-I. Photoremovable protecting groups. In *CRC Handbook of Organic Photochemistry and Photobiology*, 2nd ed.; Horspool, W., Lenci, F., Eds.; CRC Press: Boca Raton, FL, 2004, Chapter 69, pp 1–46. (b) Pelliccioli, A. P.; Wirz, J. *Photochem. Photobiol. Sci.* 2002, 1, 441–458.

(35) Rahman, M. T.; Nahar, S. K. J. Organomet. Chem. 1987, 329, 133–138.

(36) Itoh, T.; Shimizu, Y.; Kanai, M. Org. Lett. 2014, 16, 2736–2739.

(37) (a) Herrmann, A. Photochem. Photobiol. Sci. 2012, 11, 446–459.

(b) Johnson, E. C. B.; Kent, S. B. H. Chem. Commun. 2006, 1557– 1559. (c) Kim, M. S.; Diamond, S. L. Bioorg. Med. Chem. Lett. 2006, 16, 4007–4010.

(38) Sugimoto, A.; Fukuyama, T.; Sumino, Y.; Takagi, M.; Ryu, I. *Tetrahedron* **2009**, 65, 1593–1598.

(39) Wang, B.; Qin, L.; Neumann, K. D.; Uppaluri, S.; Cerny, R. L.; Dimagno, S. G. Org. Lett. 2010, 12, 3352–3355.

(40) Ito, K.; Tanaka, H.; Kayama, M. Chem. Pharm. Bull. 1977, 25, 1249–1255.

(41) Lu, J.; Guan, Z.-Z.; Gao, J.-W.; Zhang, Z.-H. Appl. Organomet. Chem. 2011, 25, 537–541.

(42) Chalker, J. M.; Wood, C. S. C.; Davis, B. G. J. Am. Chem. Soc. 2009, 131, 16346-16347.

(43) Ong-Lee, A.; Sylvester, L.; Wasley, J. W. F. J. Heterocycl. Chem. 1983, 20, 1565–1569.

(44) Augner, D.; Gerbino, D. C.; Slavov, N. Org. Lett. 2011, 13, 1629–1631.